

## O2k manual titrations: SUIT protocols with mitochondrial preparations

**OROBOROS INSTRUMENTS**, high-resolution respirometry

Schöpfstr 18, A-6020 Innsbruck, Austria

[instruments@oroboros.at](mailto:instruments@oroboros.at); [www.oroboros.at](http://www.oroboros.at)



### O2k chamber volume: 2.0 ml

Substrates	Event	Concentration in syringe (solvent)	Storage [°C]	Final conc. in 2 ml	Titration [µl]	Syringe [µl]
Pyruvate	P	2 M (H <sub>2</sub> O)	fresh	5 mM	5	25
Malate	M	0.8 M (H <sub>2</sub> O)	-20	2 mM	5	25
Glutamate	G	2 M (H <sub>2</sub> O)	-20	10 mM	10	25
Succinate*	S	1 M (H <sub>2</sub> O)	-20	10 mM	20	50
Octanoyl carnitine	Oct	0.1 M (H <sub>2</sub> O)	-20	0.2 mM	4	10
Ascorbate	As	0.8 M (H <sub>2</sub> O)	-20	2 mM	5	25
TMPD	Tm	0.2 M (H <sub>2</sub> O)	-20	0.5 mM	5	25
Cyt. c	c	4 mM (H <sub>2</sub> O)	-20	10 µM	5	25
ADP+ Mg <sup>2+</sup>	D	0.5 M (H <sub>2</sub> O)	-80	1 - 5 mM	4 - 20	25
ATP+ Mg <sup>2+</sup>	T	0.5 M (H <sub>2</sub> O)	-80	1 - 5 mM	4 - 20	25
<b>Uncoupler</b>						
CCCP <sup>‡</sup>	U	0.1 mM (EtOH)	-20	0.05 µM steps	1 µl steps	10
CCCP <sup>‡</sup>	U	1.0 mM (EtOH)	-20	0.5 µM steps	1 µl steps	10
<b>Inhibitors</b>						
Rotenone	Rot	1 mM (EtOH)	-20	0.5 µM	1	10
Malonic acid	Mna	2 M (H <sub>2</sub> O)	fresh	5 mM	5	25
Antimycin A	Ama	5 mM (EtOH)	-20	2.5 µM	1	10
Myxothiazol	Myx	1 mM (EtOH)	-20	0.5 µM	1	10
Sodium azide	Azd	4 M (H <sub>2</sub> O)	-20	≥100 mM	≥50	50
KCN	Kcn	1 M (H <sub>2</sub> O)	fresh	1.0 mM	2	10
Oligomycin	Omy	5 mM (EtOH)	-20	2.5 µM	1	10
Carboxyatractyloside	Cat	5 mM (H <sub>2</sub> O)	-20	5 µM	2	10
<b>Other</b>						
Digitonin	Dig	10 mg/ml (DMSO)	-20	10 µg·10 <sup>-6</sup> cells	1 µl 10 <sup>-6</sup>	10
Catalase in MiR06	Ctl	112,000 U/ml	-20	280 U/ml	5	25
Hydrogen peroxide for reoxygenation	Hp	200 mM	fresh		1 - 3	10

\* The concentration of S may be increased up to 50 mM after Rot to compensate for the inhibitory effect of M.

‡ 0.1 mM stock for mt-preparations with high uncoupler sensitivity; 1 mM stock for mt-preparations with low uncoupler sensitivity, intact cells in various culture media (e.g. RPMI, DMEM, EGC) and for TIP2k.

**O2k-Chamber volume: 2.0 ml**

Fluorescence probes and related	Event	Concentration in syringe (solvent)	Storage [°C]	Final conc. in 2 ml	Titration [µl]	Syringe [µl]
Amplex®UltraRed	AmR	10 mM (DMSO)	-20	10 µM	2	10
Horse radish peroxidase	HRP	500 U/ml (MiR05)	-20	1 U/ml	4	10
Superoxide dismutase	SOD	check supplier information	4-8	5 U/ml		10
Hydrogen peroxide for calibration	Hp	0.04 mM (H <sub>2</sub> O)	fresh	0.1 µM	5	10
Safranin	Saf	0.2 mM (H <sub>2</sub> O)	RT	0.25 µM	2.5	10
TMRM	Tmr	0.2 mM (H <sub>2</sub> O)	-20	0.25 µM	2.5	10
Calcium Green	CaG	2 mM (H <sub>2</sub> O)	-20	1 µM	1	10
Magnesium Green	MG	5 mM (H <sub>2</sub> O)	-20	2.5 µM	1	10

**Further abbreviations**

Atractyloside	Atr
Calcium	Ca <sup>2+</sup>
Dinitrophenol	DNP; U
Carbonyl cyanide p-trifluoromethoxyphenyl hydrazone	FCCP; U
Glucose	Glc
Glycerophosphate	Gp
Hydroxycinnamate	Hci
Oxaolacetate	Oa
Octanoate	Oca; FA
Palmitate	Paa; FA
Palmitoylcarnitine	Pal; FA
Tetraphenylphosphonium ion	TPP <sup>+</sup>

**References**

- Chinopoulos C, Kiss G, Kawamata H, Starkov AA (2014) Measurement of ADP-ATP exchange in relation to mitochondrial transmembrane potential and oxygen consumption. *Methods Enzymol* 542:333-48. »[Bioblast link](#)«
- Elustondo PA, Negoda A, Kane CL, Kane DA, Pavlov EV (2014) Spermine selectively inhibits high-conductance, but not low-conductance calcium-induced permeability transition pore. *Biochim Biophys Acta* 1847:231-40. »[Bioblast link](#)«
- Gnaiger E (2014) Mitochondrial pathways and respiratory control. An introduction to OXPHOS analysis. 4th ed. *Mitochondr Physiol Network* 19.12. OROBOROS MiPNet Publications, Innsbruck:80 pp. »[Bioblast link](#)«
- Krumschnabel G, Eigentler A, Fasching M, Gnaiger E (2014) Use of safranin for the assessment of mitochondrial membrane potential by high-resolution respirometry and fluorometry. *Methods Enzymol* 542:163-81. »[Bioblast link](#)«
- Krumschnabel G, Fontana-Ayoub M, Sumbalova Z, Heidler J, Gauper K, Fasching M, Gnaiger E (2015) Simultaneous high-resolution measurement of mitochondrial respiration and hydrogen peroxide production. *Methods Mol Biol* 1264:245-61. »[Bioblast link](#)«
- Pesta D, Gnaiger E (2012) High-resolution respirometry. OXPHOS protocols for human cells and permeabilized fibres from small biopsies of human muscle. *Methods Mol Biol* 810: 25-58. »[Bioblast link](#)«