OROBOROS O2k-Protocols Chemicals



Mitochondrial Physiology Network 14.13(06):1-4 (2016) Version 06: 2016-08-30 ©2009-2016 OROBOROS Updates: www.bioblast.at/index.php/MiPNet14.13 Medium-MiR06

Mitochondrial respiration medium -MiR06

Fasching M¹, Fontana-Ayoub M¹, Gnaiger E^{1,2}

¹ OROBOROS INSTRUMENTS Schöpfstr 18, 6020 Innsbruck, Austria Email:<u>instruments@oroboros.at</u> www.oroboros.at

²D. Swarovski Research Lab Dept of Visceral, Transplant and Thoracic Surgery Medical Univ Innsbruck, Austria www.mitofit.org



Page

- 1. Mitochondrial Respiration Medium MiR061

1. Mitochondrial respiration medium MiR06

MiR06 = MiR05 + Catalase; total volume = 1 litre (see Gnaiger et al 2000).

Compounds in MiR05/MiR06	Final conc.	FW	Addition to 1 litre final volume	Source; storage temp.
EGTA	0.5 mM	380.4	0.190 g	Sigma E 4378 (25 g); RT
MgCl₂·6 H₂O	3 mM	203.3	0.610 g	Scharlau MA 0036 (250 g); RT
Lactobionic acid	60 mM	358.3 free acid	120 ml of 0.5 M <u>K-lactobionate stock*</u>	Aldrich 153516 (100 g); RT
Taurine	20 mM	125.1	2.502 g	Sigma T 0625 (25 g); RT
KH ₂ PO ₄	10 mM	136.1	1.361 g	Merck 104873 (1 kg); RT
HEPES	20 mM	238.3	4.77 g	Sigma H 7523 (250 g); RT
D-Sucrose	110 mM	342.3	37.65 g	Roth 4621.1 or Sigma 84097** (1 kg); RT
BSA, essentially fatty acid free	1 g/l		1 g	Sigma A 6003 fraction V (25 g); 4 °C

Weigh given amounts of the listed compounds (except BSA and lactobionic acid) into a 1000 ml glass beaker, disrupt big lumps mechanically, add 800 ml H₂O and dissolve with magentic stirring at 30 °C. Add 120 ml of K-lactobionate stock solution; adjust the pH to 7.1 (5 M KOH; Sigma P 1767; 1 kg) at 30 °C. Dissolve BSA separately in subsample of the solution а (recommended to prevent foam building) and transfer back to the final solution, while stirring continuously but gently. Adjust with H_2O to a final volume of 1 litre. Check pH again and adjust if necessary. Store frozen at -20 °C in plastic vials.

* Preparation of K-lactobionate stock solution:

Dissolve 35.83 g lactobionic acid in 100 ml H_2O , adjust pH to 7.0 with KOH, adjust volume to 200 ml with H_2O .

** Sucrose: Contamination with glucose may be critical. Invert sugar (1:1 mixture of glucose and fructose) is specified for Roth 4621 (<0.04%), but not for Sigma 84097. Invert sugar content is specified for Sigma 84100 (<0.05%), however the overall specified purity for this product ($\geq 99\%$) is lower than for Sigma 8407 and Roth 4621.1 (both products $\geq 99.5\%$).

MiR06 contains the following final concentrations:

Ca ²⁺ free	0.0 µM
Mg ²⁺ free	2.1 mM
KŤ	90 mM
Na ⁺	0
EGTA free	0.46 mM
Osmolarity	330 mOsm
Ionic strength	95 mM

The ionic strength increases with the addition of substrates and adenylates, particularly in SUIT protocols with multiple substrate titrations.

EGTA A general chelator for heavy metals, with high affinity for Ca^{2+} but low affinity for Mg²⁺.

- Mg²⁺ Activation by ATP due to ATPase activity is Mg²⁺ dependent. The high quality of mitochondrial preparations cannot be tested in the absence of Mg²⁺. Physiological Mg²⁺ concentration is in the range of 1-3 mM. Several enzyme systems depend on free Mg²⁺.
- Pi The K'm is up to 1 mM in the ADP-activated OXPHOS state in heart mitochondria with glutamate/malate; 90% of maximum flux are reached at 10 mM. Is flux measurably higher at 15 mM?
- **K-lactobionate** The intracellular K⁺ concentration is high (>100 mM), adding significantly to the ionic strength. In many previous studies of isolated mitochondria, KCl was used for this reason, but a high Cl⁻ concentration is

MiPNet14.13 M	iR06
---------------	------

unphysiological and inhibitory on mitochondrial creatine kinase (and possibly on other enzymes in the intermembrane space). K-MES or K-methanesulfonate have been used successfully. Lactobionate is well established in (extracellular) organ preservation solutions (University of Wisconsin solution).

TaurineBiological membrane stabilizer and ROS scavanger. 20mM intracellular concentration in heart.

20 mM histidine was added in MiR04 (as in MiP02), as Histidine an imidazol-based buffer, with temperature dependence of pK identical to that of water (α -stat pH buffer). No effect was observed when adding histidine to MITOMED1 endothelial cells. with unpermeablized Omitted in MiR05 and MiR06 owing to increased autoxidation of TMPD and ascorbate in MiR04.

HEPES Well established buffer with pK close to 7.

Sucrose Impermeant and oxygen radical scavenger.

BSA Bovine serum albumine is a membane stabilizer, oxygen radical scavenger, and binds Ca²⁺ and free fatty acids, hence the rather expensive essentially free fatty acid free BSA is required.

- **Glutathione** No effect was observed with glutathion added to MITOMED1 with unpermeabilized or permeabilized endothelial cells (tEC); but background oxygen flux is significantly higher. This complication is avoided by not adding glutathion to the respiration medium.
- **98-03-30/04-02** Autoxidation of glutathione in MiR04 (with histidine) was variable between experiments, with $a^{\circ'}$ of -1.7 (range -2.1 to 0.3, N= 6), and $b^{\circ'}$ of 0.053, 0.065 and 0.11 in three oxygraphs (two chambers each).
- **Catalase** is an antioxidant enzyme at high intracellular activity, and improves the antioxidant quality of physiological respiration medium beyond MiR05. In the presence of high catalase activity, hydrogen peroxide, H_2O_2 , is titrated into the O2k-chamber to increase oxygen levels by up to 200 µmol/l, e.g. from 200 µM to 350 µM. MiR06 is stored like MiR05, or can be prepared by adding catalase stock solution (dissolved in MiR05) directly into the closed O2k-chamber filled with MiR05 at the start of an experiment.

Compound	Final conc.	FW	Stock solution	Addition to 2 ml final volume	Source and product code
Catalase lyophilized powder, 2,000-5,000 units/mg protein*	280 u/ml*		112000 u/ml,* dissolve in MiR05	5 µl	Sigma C9322
H ₂ O ₂		34.01	200 mM in H ₂ O, adjust to		Sigma Aldrich 516813 17.6

MiPNet14.13	MiR06 4
	pH 6 M, 50% w/w
* H ₂ O ₂	Units of enzymatic activity (u) in µmol/min; assay used by Sigma Aldrich: 'One unit will decompose 1.0 µmole of H_2O_2 per min at pH 7.0 at 25 °C, while the H_2O_2 concentration falls from 10.3 to 9.2 mM, measured by the rate of decrease of A_{240} .' Small volumes (µl) of H_2O_2 are injected into the O2k- chamber filled with MiRO6, to increase oxygen levels. A typical H_2O_2 stock concentration in the syringe is approximately 200 mM. Manual injection: Fill a 10 µl syringe with the H_2O_2 solution, inject 2-3 µl, observe the oxygen level displayed by DatLab and inject stepwise ($\Delta c_{O2} \leq 200 \ \mu$ mol/l) further H_2O_2 until the targeted oxygen level is reached. Alternatively, the "Oxystat" setup of the TIP2k may be used to reach a defined oxygen level and then maintain the oxygen concentration automatically between set limits in the
Creatine	oxystat-titration mode. MiR06Cr : MiR06Cr = MiR06 plus 20 mM creatine. MiR05Cr or MiR06Cr has to be prepared fresh every day
	from MiR05 or MiR06. Do not freeze to avoid precipitation.
Oxygen solubility	factor in MiR05 [1] at 30 °C and 37 °C is 0.92 [3]. The

Oxygen solubility factor in MiR05 [1] at 30 °C and 37 °C is 0.92 [3]. The same solubility is valid for MiR06 and MiR06Cr.

2. References

- Gnaiger E, Kuznetsov AV, Schneeberger S, Seiler R, Brandacher G, Steurer W, Margreiter R (2000) Mitochondria in the cold. In: Life in the Cold. (Heldmaier G, Klingenspor M, eds) Springer, Heidelberg, Berlin, New York, pp. 431-42.
- Rasmussen HN, Rasmussen UF (2003) Oxygen solubilities of media used in electrochemical respiration measurements. Analyt Biochem 319: 105-13. *The manuscript* (Gnaiger et al 2000) was sent to Dr. H. Rasmussen, and we appreciate that the oxygen solubility of MiR05 was then determined and published (Rasmussen 2003). *Surprisingly, no reference was made in ref.* (Rasmussen et al 2003) to the original publication on MiR05 (MiPNet08.05).

MiPNet03.02

Selected media and chemicals for respirometry with mitochondria and permeabilized cells.



Acknowledgements

Contribution to K-Regio project *MitoCom Tyrol*, funded in part by the Tyrolian Government and the European Regional Development Fund (ERDF). <u>http://www.bioblast.at/index.php/MitoCom O2k-Fluorometer</u>

Author contributions

Gnaiger E with collaboration of Kuznetsov AV was responsible for the development and testing of MiR05 (Gnaiger et al 2000). All authors contributed to various details in the development of MiR06.

Supplementary material:

http://wiki.oroboros.at/index.php/MiPNet14.13 Medium-MiR06